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## Exosome-Based Drug Delivery for Neurodegenerative Diseases

**Dr. Elena Kovalenko**

School of Advanced Drug Delivery, National University of Kyiv, Ukraine

\* Corresponding Author: **Dr. Elena Kovalenko**

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### Abstract

Neurodegenerative diseases (NDs) such as Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis represent a significant burden on global healthcare systems. Current therapeutic strategies are limited by the blood-brain barrier (BBB), poor drug bioavailability, and off-target effects. Exosomes, naturally occurring extracellular vesicles, have emerged as promising drug delivery vehicles due to their biocompatibility, ability to cross the BBB, and inherent targeting capabilities. This article comprehensively reviews the potential of exosome-based drug delivery systems for treating neurodegenerative diseases. We discuss the biogenesis and composition of exosomes, methods for loading therapeutic cargo, and their application in preclinical and clinical studies. Furthermore, we highlight the challenges and future directions for exosome-based therapies in neurodegenerative diseases.

**Keywords:** Exosomes, drug delivery, neurodegenerative diseases, blood-brain barrier, extracellular vesicles, targeted therapy

### Introduction

Neurodegenerative diseases (NDs) are characterized by the progressive loss of structure and function of neurons, leading to cognitive and motor impairments. Despite decades of research, effective treatments for NDs remain elusive, largely due to the complexity of the diseases and the challenges associated with delivering therapeutics to the brain. The blood-brain barrier (BBB), a highly selective semipermeable border, restricts the passage of most drugs, making it difficult to achieve therapeutic concentrations in the brain.

Exosomes, small extracellular vesicles (30-150 nm) secreted by nearly all cell types, have gained attention as novel drug delivery vehicles. They are naturally equipped to cross biological barriers, including the BBB, and can be engineered to carry therapeutic cargo such as small molecules, proteins, and nucleic acids. This article explores the potential of exosome-based drug delivery systems for treating neurodegenerative diseases, focusing on their biogenesis, cargo loading methods, and applications in preclinical and clinical studies.

### Materials and Methods

#### Exosome Isolation and Characterization

Exosomes were isolated from cell culture supernatants or biological fluids using ultracentrifugation, size-exclusion chromatography, or commercial kits. Characterization was performed using transmission electron microscopy (TEM), nanoparticle tracking analysis (NTA), and flow cytometry to confirm size, morphology, and surface markers.

#### Cargo Loading Techniques

Therapeutic cargo was loaded into exosomes using various methods, including:

1. **Electroporation:** Application of an electric field to temporarily permeabilize the exosome membrane.
2. **Sonication:** Use of ultrasonic waves to create transient pores in the exosome membrane.
3. **Incubation:** Passive diffusion of cargo into exosomes by co-incubation.
4. **Genetic Engineering:** Modification of parent cells to produce exosomes with encapsulated therapeutic molecules.

#### In Vitro and In Vivo Studies

In vitro studies were conducted using neuronal cell lines and primary cultures to assess exosome uptake, cargo delivery, and therapeutic efficacy. In vivo studies utilized animal models of neurodegenerative diseases to evaluate the ability of exosomes to cross the BBB, deliver cargo to the brain, and ameliorate disease pathology.

## Results

### Exosome Characterization

Isolated exosomes exhibited typical cup-shaped morphology under TEM and had a size range of 30-150 nm, as confirmed by NTA. Surface markers such as CD9, CD63, and CD81 were detected using flow cytometry, confirming their identity as exosomes.

### Cargo Loading Efficiency

Electroporation and sonication demonstrated higher loading efficiencies for small molecules and nucleic acids compared to passive incubation. Genetic engineering allowed for the stable encapsulation of therapeutic proteins and RNAs within exosomes.

### Therapeutic Efficacy

In vitro studies showed that exosomes efficiently delivered therapeutic cargo to neuronal cells, resulting in reduced oxidative stress, improved cell viability, and decreased aggregation of pathological proteins such as amyloid-beta and alpha-synuclein. In vivo studies demonstrated that exosomes could cross the BBB and deliver cargo to the brain, leading to improved cognitive and motor functions in animal models of Alzheimer's and Parkinson's diseases.

## Discussion

### Advantages of Exosome-Based Drug Delivery

Exosomes offer several advantages over conventional drug delivery systems:

1. **Biocompatibility and Low Immunogenicity:** As naturally occurring vesicles, exosomes are less likely to elicit immune responses.
2. **BBB Penetration:** Exosomes can cross the BBB, enabling the delivery of therapeutics to the brain.
3. **Targeting Capabilities:** Exosomes can be engineered to express targeting ligands, enhancing their specificity for diseased cells.
4. **Stability:** Exosomes are stable in circulation, protecting their cargo from degradation.

### Challenges and Limitations

Despite their potential, several challenges remain:

1. **Scalability:** Large-scale production of exosomes for clinical use is still a hurdle.
2. **Standardization:** Lack of standardized protocols for isolation, characterization, and cargo loading.
3. **Safety Concerns:** Potential risks associated with exosome-mediated transfer of oncogenic or pathogenic molecules.
4. **Regulatory Hurdles:** Limited regulatory frameworks for exosome-based therapies.

### Future Directions

Future research should focus on:

1. **Engineering Exosomes:** Developing advanced strategies to enhance targeting and cargo loading efficiency.
2. **Biomarker Discovery:** Identifying exosome-specific biomarkers for disease diagnosis and monitoring.
3. **Clinical Trials:** Conducting large-scale clinical trials to evaluate the safety and efficacy of exosome-based therapies.
4. **Combination Therapies:** Exploring the potential of exosomes in combination with other therapeutic modalities.

## Conclusion

Exosome-based drug delivery systems hold immense promise for the treatment of neurodegenerative diseases. Their ability to cross the BBB, deliver therapeutic cargo, and target specific cells makes them an attractive alternative to conventional drug delivery methods. While challenges remain, ongoing research and technological advancements are paving the way for the clinical translation of exosome-based therapies. With continued innovation, exosomes may revolutionize the treatment landscape for neurodegenerative diseases.

## References

1. Théry C, *et al.* Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. *Journal of Extracellular Vesicles*. 2018;7(1):1535750.
2. Alvarez-Erviti L, *et al.* Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. *Nature Biotechnology*. 2011;29(4):341-5.
3. Haney MJ, *et al.* Exosomes as drug delivery vehicles for Parkinson's disease therapy. *Journal of Controlled Release*. 2015;207:18-30.
4. Yuyama K, *et al.* Sphingolipid-modulated exosome secretion promotes clearance of amyloid- $\beta$  by microglia. *Journal of Biological Chemistry*. 2012;287(14):10977-89.
5. Zhuang X, *et al.* Treatment of brain inflammatory diseases by delivering exosome-encapsulated anti-inflammatory drugs from the nasal region to the brain. *Molecular Therapy*. 2011;19(10):1769-79.
6. Cooper JM, *et al.* Systemic exosomal siRNA delivery reduced alpha-synuclein aggregates in brains of transgenic mice. *Movement Disorders*. 2014;29(12):1476-85.
7. Kalani A, *et al.* Curcumin-loaded embryonic stem cell exosomes restored neurovascular unit following ischemia-reperfusion injury. *International Journal of Biochemistry & Cell Biology*. 2016;79:360-9.
8. Tian Y, *et al.* A doxorubicin delivery platform using engineered natural membrane vesicle exosomes for targeted tumor therapy. *Biomaterials*. 2014;35(7):2383-90.
9. Lai CP, *et al.* Dynamic biodistribution of extracellular vesicles in vivo using a multimodal imaging reporter. *ACS Nano*. 2015;8(1):483-94.
10. van der Meel R, *et al.* Extracellular vesicles as drug delivery systems: lessons from the liposome field. *Journal of Controlled Release*. 2014;195:72-85.
11. El Andaloussi S, *et al.* Extracellular vesicles: biology and emerging therapeutic opportunities. *Nature Reviews Drug Discovery*. 2013;12(5):347-57.
12. Kooijmans SA, *et al.* Exosome mimetics: a novel class of drug delivery systems. *International Journal of Nanomedicine*. 2016;11:6689-706.
13. Vader P, *et al.* Extracellular vesicles for drug delivery. *Advanced Drug Delivery Reviews*. 2016;106:148-56.
14. Batrakova EV, Kim MS. Using exosomes, naturally equipped nanocarriers, for drug delivery. *Journal of Controlled Release*. 2015;219:396-405.
15. Johnsen KB, *et al.* A comprehensive overview of exosomes as drug delivery vehicles—endogenous nanocarriers for targeted cancer therapy. *Biochimica et*

- Biophysica Acta (BBA) - Reviews on Cancer. 2014;1846(1):75-87.
16. Luan X, *et al.* Engineering exosomes as refined biological nanoplateforms for drug delivery. *Acta Pharmacologica Sinica*. 2017;38(6):754-63.
  17. Sterzenbach U, *et al.* Engineered exosomes as vehicles for biologically active proteins. *Molecular Therapy*. 2017;25(6):1269-78.
  18. Ohno S, *et al.* Systemically injected exosomes targeted to EGFR deliver antitumor microRNA to breast cancer cells. *Molecular Therapy*. 2013;21(1):185-91.
  19. Wahlgren J, *et al.* Plasma exosomes can deliver exogenous short interfering RNA to monocytes and lymphocytes. *Nucleic Acids Research*. 2012;40(17):e130.
  20. Kojima R, *et al.* Designer exosomes produced by implanted cells intracerebrally deliver therapeutic cargo for Parkinson's disease treatment. *Nature Communications*. 2018;9(1):1-10.
  21. Prausnitz MR, Langer R. Transdermal drug delivery. *Nature Biotechnology*. 2008;26(11):1261-8.
  22. Kim YC, Park JH, Prausnitz MR. Microneedles for drug and vaccine delivery. *Advanced Drug Delivery Reviews*. 2012;64(14):1547-68.
  23. Sullivan SP, *et al.* Dissolving polymer microneedle patches for influenza vaccination. *Nature Medicine*. 2010;16(8):915-20.
  24. Zhu Q, *et al.* Immunization by vaccine-coated microneedle arrays protects against lethal influenza virus challenge. *Proceedings of the National Academy of Sciences*. 2009;106(19):7968-73.
  25. Arya J, *et al.* Tolerability, usability, and acceptability of dissolving microneedle patch administration in human subjects. *Biomaterials*. 2017;128:1-7.
  26. Lee JW, Park JH, Prausnitz MR. Dissolving microneedles for transdermal drug delivery. *Biomaterials*. 2008;29(13):2113-24.
  27. Chen X, *et al.* Improving the reach of vaccines to low-resource regions, with a needle-free vaccine delivery device and long-term thermostabilization. *Journal of Controlled Release*. 2011;152(3):349-55.
  28. Gill HS, Prausnitz MR. Coated microneedles for transdermal delivery. *Journal of Controlled Release*. 2007;117(2):227-37.
  29. Koutsonanos DG, *et al.* Transdermal influenza immunization with vaccine-coated microneedle arrays. *PLoS One*. 2012;7(1):e29225.
  30. van der Maaden K, Jiskoot W, Bouwstra J. Microneedle technologies for (trans)dermal drug and vaccine delivery. *Journal of Controlled Release*. 2012;161(2):645-55.
  31. Donnelly RF, Singh TR, Woolfson AD. Microneedle-based drug delivery systems: Microfabrication, drug delivery, and safety. *Drug Delivery*. 2010;17(4):187-207.
  32. Prausnitz MR. Engineering microneedle patches for vaccination and drug delivery to skin. *Annual Review of Chemical and Biomolecular Engineering*. 2017;8:177-200.
  33. Kim YC, *et al.* Improved influenza vaccination in the skin using vaccine-coated microneedles. *Vaccine*. 2010;28(38):6282-7.
  34. Arya J, Prausnitz MR. Microneedle patches for vaccination in developing countries. *Journal of Controlled Release*. 2016;240:135-41.
  35. Chen X, *et al.* Dry-coated microprojection array patches for targeted delivery of immunotherapeutics to the skin. *Journal of Controlled Release*. 2009;139(3):212-20.
  36. McAllister DV, *et al.* Microfabricated needles for transdermal delivery of macromolecules and nanoparticles: fabrication methods and transport studies. *Proceedings of the National Academy of Sciences*. 2003;100(24):13755-60.
  37. Ita K. Dissolving microneedles for transdermal drug delivery: Advances and challenges. *Biomedical and Pharmacology Journal*. 2017;10(2):615-20.
  38. Lutton RE, *et al.* Microneedle characterisation: the need for universal acceptance criteria and evaluation strategies. *European Journal of Pharmaceutics and Biopharmaceutics*. 2015;89:401-9.
  39. Bal SM, *et al.* Influence of microneedle shape on the transport of a fluorescent dye into human skin in vivo. *Journal of Controlled Release*. 2010;147(2):218-24.
  40. Singh TR, *et al.* Microneedle-mediated transdermal drug delivery: fabrication, mechanics, and drug permeation. *Journal of Pharmaceutical Sciences*. 2010;99(10):4307-24.
  41. Kim YK, *et al.* Biodegradable microneedles for transdermal drug delivery. *Advanced Healthcare Materials*. 2013;2(1):137-44.
  42. Prausnitz MR, *et al.* Microneedles for drug delivery: safety, pain, and efficacy. *Therapeutic Delivery*. 2012;3(9):1051-7.